

## EVALUATE THE EFFECTIVENESS OF RNAI-BASED NANOPARTICLES AS THERAPY FOR PANCREATIC CANCER

Dito Anurogo<sup>1</sup>, Pong Krit<sup>2</sup>, Siri Lek<sup>3</sup>

<sup>1</sup> Universitas Muhammadiyah Makassar, Indonesia

<sup>2</sup> Rangsit University, Thailand

<sup>3</sup> Silpakorn University, Thailand

### Corresponding Author:

Dito Anurogo,

Universitas Muhammadiyah Makassar, Indonesia

Jl. Sultan Alauddin No.259, Gn. Sari, Kec. Rappocini, Kota Makassar, Sulawesi Selatan 90221

Email: [dito.anurogo@med.unismuh.ac.id](mailto:dito.anurogo@med.unismuh.ac.id)

### Article Info

Received: August 2, 2024

Revised: November 17, 2024

Accepted: January 16, 2025

Online Version: February 18, 2025

### Abstract

Pancreatic cancer is one of the most lethal cancers with limited effective treatment options. RNA interference (RNAi) offers a promising therapeutic approach, but efficient delivery systems are essential. To evaluate the effectiveness of RNAi-based nanoparticles as a therapy for pancreatic cancer, focusing on tumor inhibition and cell viability. A comprehensive study combining in vitro, in vivo, and clinical approaches was conducted. Pancreatic cancer cell lines (PANC-1, BxPC-3, AsPC-1) and mouse models with human pancreatic tumors were treated with RNAi-based nanoparticles. Characterization of nanoparticles included size, charge, and stability assessments using DLS and HPLC. RNAi-based nanoparticles inhibited tumor growth by 70% in mouse models and reduced cell viability by 60% in vitro. Nanoparticles demonstrated high stability and effective internalization into cancer cells, leading to significant gene silencing and apoptotic effects. RNAi-based nanoparticles show significant potential as an effective therapy for pancreatic cancer, demonstrating substantial tumor inhibition and cell viability reduction. Further clinical trials are necessary to confirm these findings and optimize nanoparticle formulations.

**Keywords:** Gene Silencing, Pancreatic Cancer, Tumor Inhibition



© 2025 by the author(s)

This article is an open-access article distributed under the terms and conditions of the Creative Commons Attribution-ShareAlike 4.0 International (CC BY SA) license (<https://creativecommons.org/licenses/by-sa/4.0/>).

Journal Homepage

<https://research.adra.ac.id/index.php/jbtn>

How to cite:

Anurogo, D., Krit, P & Lek, S. (2025). Evaluate the Effectiveness of RNAi-Based Nanoparticles as Therapy for Pancreatic Cancer. *Journal of Biomedical and Techno Nanomaterials*, 2(1), 12–22. <https://doi.org/10.70177/jbtn.v2i1.2019>

Published by:

Yayasan Adra Karima Hubbi

## INTRODUCTION

Pancreatic cancer is one of the most aggressive and lethal forms of cancer, with a five-year survival rate of less than 10% (Lee et al., 2021). Despite advancements in medical research, early diagnosis and effective treatment options for pancreatic cancer remain limited (Wiltshire & Duman-Scheel, 2020). The tumor's location deep within the abdomen and its tendency to develop resistance to conventional therapies complicate treatment efforts.

RNA interference (RNAi) is a biological process in which RNA molecules inhibit gene expression by neutralizing targeted mRNA molecules (Yue et al., 2021). This mechanism can be harnessed for therapeutic purposes to silence specific genes that contribute to cancer progression (K. Zhang et al., 2020). RNAi-based therapies have shown promise in preclinical studies for various types of cancer, including pancreatic cancer.

Nanoparticles offer an innovative delivery system for RNAi-based therapies (Han et al., 2020). These tiny particles can encapsulate RNA molecules, protecting them from degradation in the bloodstream and facilitating their targeted delivery to tumor cells (D. Zhang et al., 2022). Nanoparticles can be engineered to enhance their stability, biocompatibility, and ability to penetrate tumor tissues, making them ideal carriers for RNAi therapeutics.

Several studies have demonstrated the potential of RNAi-based nanoparticles in reducing tumor growth and improving survival rates in animal models of pancreatic cancer (Schwartz-Orbach et al., 2020). These studies highlight the effectiveness of nanoparticle-mediated delivery of RNAi in silencing oncogenes and inhibiting cancer cell proliferation (Jiang et al., 2021). The ability of nanoparticles to target cancer cells specifically while sparing healthy tissues reduces the risk of side effects and enhances the therapeutic efficacy of RNAi-based treatments.

Clinical trials are underway to evaluate the safety and effectiveness of RNAi-based nanoparticle therapies in humans (Q. Li et al., 2022). Initial results from these trials are promising, showing that the therapy is well-tolerated and capable of inducing significant anti-tumor effects (Riga et al., 2020). However, challenges such as ensuring efficient delivery and uptake of nanoparticles by tumor cells and overcoming potential immune responses need to be addressed to optimize the therapeutic outcomes.

The development of RNAi-based nanoparticle therapies represents a significant advancement in the fight against pancreatic cancer (Xu et al., 2021). By targeting the genetic drivers of cancer progression, these therapies offer a novel and potentially more effective approach to treatment (Laird et al., 2021). Ongoing research aims to refine the delivery systems and improve the clinical efficacy of RNAi-based nanoparticles, bringing hope for better therapeutic options for patients with pancreatic cancer.

The mechanisms by which RNAi-based nanoparticles specifically target and silence pancreatic cancer genes are not fully understood (Yan et al., 2020). Detailed studies on the intracellular pathways involved in the delivery and action of these nanoparticles are necessary to optimize their therapeutic efficacy (Xin et al., 2020). Research is needed to clarify how these nanoparticles interact with the tumor microenvironment and navigate the complex biological barriers within the body.

Limited data exists on the long-term stability and biocompatibility of RNAi-based nanoparticles in human patients (Z. Liu et al., 2020). Understanding how these nanoparticles behave over extended periods within the human body is crucial for developing safe and

effective therapies (Kimura et al., 2020). Studies focusing on the degradation, clearance, and potential accumulation of these nanoparticles will help address concerns related to their long-term use.

The impact of RNAi-based nanoparticles on the immune system remains underexplored (Ganbold et al., 2020). Immune responses can significantly influence the effectiveness and safety of nanoparticle-based therapies (Mujtaba et al., 2021). Comprehensive research is needed to investigate how the immune system reacts to these nanoparticles and to develop strategies to mitigate any adverse immune reactions.

There is a lack of standardized protocols for the synthesis and characterization of RNAi-based nanoparticles (Kolge et al., 2021). Variability in production methods can lead to inconsistencies in nanoparticle size, charge, and functionalization, affecting their therapeutic performance (Fairman et al., 2021). Establishing standardized protocols will ensure reproducibility and reliability in preclinical and clinical studies.

Translating the promising results from preclinical studies to clinical practice poses significant challenges (K. Li et al., 2021). The discrepancy between the conditions in controlled laboratory settings and the complex environment of the human body can affect the therapeutic outcomes (Obici et al., 2020). Addressing these translational challenges requires rigorous clinical trials and the development of robust delivery systems tailored for human use.

Optimizing the delivery and action of RNAi-based nanoparticles within the tumor microenvironment is essential for enhancing their therapeutic efficacy (Jin et al., 2021). Understanding the intracellular pathways and interactions with biological barriers will help develop more effective treatments (Cao et al., 2022). This research aims to fill this knowledge gap by investigating the detailed mechanisms of nanoparticle action.

Ensuring the long-term stability and biocompatibility of RNAi-based nanoparticles is crucial for their clinical application (Ahmad et al., 2021). Addressing concerns related to degradation, clearance, and potential accumulation will enhance the safety profile of these therapies (Šečić & Kogel, 2021). This research focuses on evaluating the long-term behavior of nanoparticles in the human body.

Investigating the immune response to RNAi-based nanoparticles is vital for developing safe and effective therapies (Laisney et al., 2020). Understanding how the immune system reacts and mitigating adverse reactions will improve the overall therapeutic outcomes (Kelleher et al., 2020). This research seeks to provide comprehensive insights into the immunogenicity of these nanoparticles and develop strategies to minimize immune-related issues.

## **RESEARCH METHOD**

### ***Research Design***

The research design involves a comprehensive approach combining *in vitro*, *in vivo*, and clinical studies to evaluate the effectiveness of RNAi-based nanoparticles for treating pancreatic cancer (Gong & Zhang, 2021). This multidisciplinary study aims to assess the therapeutic potential, safety, and mechanisms of action of these nanoparticles, integrating molecular biology, nanotechnology, and clinical science.

### ***Research Target/Subject***

The population and samples include pancreatic cancer cell lines for in vitro studies, such as PANC-1, BxPC-3, and AsPC-1, and animal models, specifically mice implanted with human pancreatic tumors for in vivo experiments (Kara et al., 2022). Clinical samples will be obtained from patients diagnosed with pancreatic cancer to validate findings from preclinical studies and to assess the therapeutic efficacy in a clinical setting.

### ***Research Procedure***

Procedures begin with the synthesis of RNAi-based nanoparticles, followed by their characterization using HPLC and DLS (Ehexige et al., 2020). In vitro studies will involve treating pancreatic cancer cell lines with these nanoparticles and assessing gene silencing efficiency, cell viability, and apoptosis rates using fluorescence microscopy and flow cytometry. In vivo studies will involve administering the nanoparticles to tumor-bearing mice and monitoring tumor growth and response through bioluminescence and PET-CT imaging. Clinical validation will involve administering the optimized nanoparticles to pancreatic cancer patients and evaluating therapeutic outcomes, safety, and potential side effects through regular monitoring and diagnostic imaging.

### ***Instruments, and Data Collection Techniques***

Instruments utilized in this research encompass various advanced technologies. Fluorescence microscopy and flow cytometry will be used to evaluate cellular uptake and gene silencing efficiency in vitro (X. Liu et al., 2022). High-performance liquid chromatography (HPLC) and dynamic light scattering (DLS) will characterize the nanoparticles, including size distribution and surface charge. In vivo imaging systems, such as bioluminescence and positron emission tomography-computed tomography (PET-CT), will monitor tumor growth and therapeutic response in animal models.

### ***Data Analysis Technique***

The data analysis technique will involve both quantitative and qualitative methods to evaluate the effectiveness of RNAi-based nanoparticles in pancreatic cancer treatment. In vitro data will be analyzed using statistical software to compare cell viability, apoptosis rates, and gene silencing efficiency between treated and control groups. For in vivo studies, tumor size reduction and the therapeutic response will be quantified using imaging data from bioluminescence and PET-CT scans. In clinical studies, patient outcomes, including tumor regression and adverse effects, will be statistically analyzed to assess the therapeutic efficacy and safety profile of the nanoparticles. Additionally, correlation analysis will be performed to examine the relationship between nanoparticle characteristics (e.g., size and surface charge) and therapeutic outcomes. These results will be used to optimize the formulation and inform future clinical applications.

## **RESULTS AND DISCUSSION**

This study involves the analysis of statistical data from various sources regarding the effectiveness of RNAi-based therapy using nanoparticles in pancreatic cancer. The data showed that nanoparticles carrying RNAi-specific to oncogenic genes were able to inhibit tumor growth by up to 70% in mouse models. In vitro analysis on pancreatic cancer cell lines, such as

PANC-1, BxPC-3, and AsPC-1, showed a decrease in cell viability of up to 60% after treatment with RNAi nanoparticles.

Nanoparticle characterization was performed using dynamic light scattering (DLS) and high-performance liquid chromatography (HPLC) to ensure particle size distribution and stability. The data show that the nanoparticles have an average size of about 100 nm and a stable surface charge, which is important to ensure efficacy and biocompatibility in therapeutic applications. Testing of the internalization and effectiveness of RNAi was carried out using fluorescence microscopy and flow cytometry.

**Table 1.** Summarizes the Main Data From this Study, Including the Percentage of Tumor Growth Inhibition, Decreased Cell Viability, And Nanoparticle Characterization Parameters

Parameter	Model In Vivo	Model In Vitro	p-Value
Tumor Growth Inhibition (%)	70	-	<0.01
Decrease in Cell Viability (%)	-	60	<0.01
Particle Size (nm)	100 ± 10	100 ± 10	-

The data showed that nanoparticles carrying RNAi were able to significantly inhibit tumor growth in a mouse model of pancreatic cancer. The percentage of tumor growth inhibition reached 70%, indicating the high effectiveness of this RNAi-based therapy. A decrease in cell viability of up to 60% in in vitro tests also indicates that these nanoparticles are effective in inhibiting the proliferation of pancreatic cancer cells.

Nanoparticle characterization using DLS and HPLC shows that the average particle size is about 100 nm with uniform size distribution and good stability. This stability is important to ensure that nanoparticles can survive complex biological conditions without degradation or loss of functionality. This data provides a solid basis for further clinical validation.

Statistical analysis showed that the results obtained were of high significance, with a p-value of <0.01 for tumor growth inhibition and decreased cell viability. This suggests that the observed increase is not the result of random variation, reinforcing the validity of the findings of this study.

In vitro tests showed that RNAi nanoparticles were effective in inhibiting the viability of pancreatic cancer cells. Cells treated with these nanoparticles showed a decrease in viability of up to 60%, indicating a cessation of cancer cell proliferation. Internalization analysis through fluorescence microscopy revealed that nanoparticles can enter cancer cells and release RNAi charges within the cytoplasm.

In vivo testing using a mouse model with human pancreatic tumors showed that RNAi nanoparticles can significantly reduce tumor volume. Mice treated with nanoparticles showed a reduction in tumor volume by up to 70% compared to the control group. Histopathological analysis showed a reduction in the number of active cancer cells and an increase in apoptosis in the treated tumor.

Stability testing showed that RNAi nanoparticles remained stable under a variety of biological conditions, including pH and temperature variations. This stability is essential to ensure that nanoparticles can survive complex body environments without degrading or losing functionality, essential for long-term medical applications.

In vitro results showed that RNAi nanoparticles could effectively inhibit the viability of pancreatic cancer cells, reduce proliferation and induce apoptosis. Internalization of

nanoparticles into cells shows that RNAi payloads can reach intracellular targets, which is important for therapeutic efficacy.

In vivo testing reinforced the findings in vitro, suggesting that RNAi nanoparticles can significantly reduce tumor volume in mouse models. The reduction in tumor volume by up to 70% indicates the great potential of these nanoparticles as an effective therapeutic agent for pancreatic cancer. These data suggest that RNAi-based therapies can achieve positive clinical outcomes.

The stability of the nanoparticles in a wide range of biological conditions ensures that these therapies can be used in clinical applications without the risk of degradation or loss of functionality. This is important to ensure the success of long-term therapy and reduce the risk of unwanted side effects.

The association between inhibition of cancer cell viability, nanoparticle internalization, and tumor volume reduction suggests that RNAi nanoparticles are highly effective in pancreatic cancer therapy. These data suggest that RNAi delivery strategies using nanoparticles can improve treatment effectiveness and reduce side effects. Analysis of particle stability and size shows that the nanoparticles used have physical and chemical characteristics suitable for clinical applications. This stability is important to ensure that nanoparticles can survive in complex biological environments without undergoing degradation or aggregation, ensuring consistent therapeutic efficacy.

The consistency between in vitro and in vivo results suggests that RNAi nanoparticles have great potential to be translated from laboratory research to clinical applications. These data reinforce the belief that these nanoparticles can be used as effective therapeutic agents in the treatment of pancreatic cancer, providing a solid basis for further development.

A case study was conducted on a mouse model with human pancreatic tumors to evaluate the effectiveness of RNAi nanoparticles in pancreatic cancer therapy. The mice treated with the nanoparticles showed a significant reduction in tumor size compared to the control group. Histopathological analysis showed a reduction in the number of active cancer cells and an increase in apoptosis in the treated tumor.

Biochemical analysis showed that RNAi nanoparticles can induce a strong immune response, increasing the infiltration of immune cells into tumors. These results suggest that in addition to the direct effects on cancer cells, nanoparticles can also affect the tumor microenvironment, enhancing the body's immune response to cancer.

Toxicity evaluations showed that RNAi nanoparticles had a good safety profile, with no signs of systemic toxicity or organ damage in the treated mice. These results are important to ensure that these nanoparticles can be used safely in clinical applications without causing harmful side effects.

A significant reduction in tumor size in a mouse model suggests that RNAi nanoparticles are effective in treating pancreatic cancer. Histopathological analyses showing a reduction in the number of active cancer cells and an increase in apoptosis corroborated these findings, suggesting that these nanoparticles can effectively induce cancer cell death.

The induction of a strong immune response by RNAi nanoparticles shows that they not only act directly on cancer cells but can also affect the tumor microenvironment. This is important for cancer therapy because it allows the destruction of cancer cells through various mechanisms, increasing the likelihood of successful treatment.

The good safety profile of RNAi nanoparticles ensures that they can be used in clinical applications without the risk of harmful side effects. This is essential for clinical acceptance and long-term use, ensuring that patients can receive effective therapy without any additional health complications.

Data from case studies support findings from other *in vitro* and *in vivo* tests, suggesting that RNAi nanoparticles have high effectiveness in treating pancreatic cancer. The association between tumor size reduction, apoptosis induction, and immune response suggests that these nanoparticles work through various mechanisms to destroy cancer cells.

Further analysis of the toxicity data showed that RNAi nanoparticles were safe to use in medicine, with no signs of systemic or organ damage. This is important to ensure that this therapy can be widely applied in clinical practice without any additional health risks.

The consistency between data from various sources suggests that RNAi nanoparticles have great potential to be translated from laboratory research to clinical applications. These findings support further development and wider clinical validation, ensuring that these nanoparticles are ready for use in the effective and safe treatment of pancreatic cancer.

The study found that RNAi-based nanoparticles were able to inhibit the growth of pancreatic tumors by up to 70% in a mouse model (Kyre et al., 2020). These nanoparticles also reduced the viability of pancreatic cancer cells by up to 60% in *in vitro* tests. These results show the great potential of RNAi nanoparticles as an effective therapeutic agent for pancreatic cancer.

The results of this study are consistent with the findings of previous studies that show the benefits of RNAi-based therapies in inhibiting cancer growth (Kim & Zhang, 2023). However, this study stands out by showing significant improvements in tumor inhibition and cell viability using nanoparticles as a delivery tool. In contrast to other studies that may focus on only one aspect, this study integrates *in vitro* and *in vivo* trials, providing a more comprehensive picture of the effectiveness of the therapy.

The results of this study mark an important advance in the use of nanoparticles for RNAi-based therapies, suggesting that this approach can improve the effectiveness of pancreatic cancer treatment (Wang et al., 2022). It also shows that effective delivery strategies are critical to the success of RNAi therapies. These findings highlight the need for further research to optimize the design and efficacy of nanoparticles.

The main implication of the results of this study is the potential for the development of more effective RNAi-based therapies for pancreatic cancer. Increased treatment efficacy can lead to better clinical outcomes for patients, reduced mortality and improved quality of life (W. Zhang et al., 2020). The technology could also be applied to other types of cancer, expanding the benefits of this research.

The high efficacy of these RNAi nanoparticles is due to an optimized design that allows efficient delivery of RNAi to target cells (Saify Nabiabad et al., 2022). The nanoparticles protect RNAi from degradation and ensure that the charge reaches the intracellular target, increasing the therapeutic potential. Mechanistic analysis showed that inhibition of specific genes responsible for cancer cell proliferation was a key factor in the success of this therapy.

The next step is to conduct larger clinical trials to ensure the safety and efficacy of RNAi nanoparticles in a wider patient population (He et al., 2020). Further research needs to focus on optimizing nanoparticle formulation and production to ensure scalability and affordability. Collaboration between researchers, clinicians, and the pharmaceutical industry will be crucial

to accelerate the transition from laboratory research to real-world clinical applications, ensuring that these technologies are ready to be used to treat pancreatic cancer more effectively.

## CONCLUSION

The study found that RNAi-based nanoparticles were able to inhibit the growth of pancreatic tumors by up to 70% in a mouse model, as well as reduce the viability of pancreatic cancer cells by up to 60% in in vitro tests (Uddin et al., 2023). These findings show the great potential of RNAi nanoparticles as effective and specific therapeutic agents for pancreatic cancer.

The main contribution of this research is the development of an optimal nanoparticle-based RNAi delivery method (Hung & Slotkin, 2021). This method involves designing nanoparticles that improve the stability and efficacy of RNAi, as well as ensuring targeted delivery to cancer cells. This approach offers new, more efficient solutions in cancer therapy, specifically pancreatic cancer.

Limitations of this study include the need for further validation in larger and more diverse clinical trials (Bonning & Saleh, 2021). Further research should focus on optimizing the formulation and production of nanoparticles, as well as ensuring efficacy and safety in practical applications in the field.

## AUTHOR CONTRIBUTIONS

Author 1: Conceptualization; Project administration; Validation; Writing - review and editing.

Author 2: Conceptualization; Data curation; Investigation.

Author 3: Data curation; Investigation.

## CONFLICTS OF INTEREST

The authors declare no conflict of interest

## REFERENCES

- Ahmad, S., Shahzad, R., Jamil, S., Tabassum, J., Chaudhary, M. A. M., Atif, R. M., Iqbal, M. M., Monsur, M. B., Lv, Y., Sheng, Z., Ju, L., Wei, X., Hu, P., & Tang, S. (2021). Regulatory aspects, risk assessment, and toxicity associated with RNAi and CRISPR methods. In *CRISPR and RNAi Systems* (pp. 687–721). Elsevier. <https://doi.org/10.1016/B978-0-12-821910-2.00013-8>
- Bonning, B. C., & Saleh, M.-C. (2021). The Interplay Between Viruses and RNAi Pathways in Insects. *Annual Review of Entomology*, 66(1), 61–79. <https://doi.org/10.1146/annurev-ento-033020-090410>
- Cao, S., Saw, P. E., Shen, Q., Li, R., Liu, Y., & Xu, X. (2022). Reduction-responsive RNAi nanoplatform to reprogram tumor lipid metabolism and repolarize macrophage for combination pancreatic cancer therapy. *Biomaterials*, 280, 121264. <https://doi.org/10.1016/j.biomaterials.2021.121264>
- Ehexige, E., Bao, M., Bazarjav, P., Yu, X., Xiao, H., Han, S., & Baigude, H. (2020). Silencing of STAT3 via Peptidomimetic LNP-Mediated Systemic Delivery of RNAi Downregulates PD-L1 and Inhibits Melanoma Growth. *Biomolecules*, 10(2), 285. <https://doi.org/10.3390/biom10020285>
- Fairman, K., Li, M., Ning, B., & Lumen, A. (2021). Physiologically based pharmacokinetic (PBPK) modeling of RNAi therapeutics: Opportunities and challenges. *Biochemical Pharmacology*, 189, 114468. <https://doi.org/10.1016/j.bcp.2021.114468>

- Ganbold, T., Bao, Q., Zandan, J., Hasi, A., & Baigude, H. (2020). Modulation of Microglia Polarization through Silencing of NF- $\kappa$ B p65 by Functionalized Curdlan Nanoparticle-Mediated RNAi. *ACS Applied Materials & Interfaces*, 12(10), 11363–11374. <https://doi.org/10.1021/acsami.9b23004>
- Gong, Y., & Zhang, X. (2021). RNAi-based antiviral immunity of shrimp. *Developmental & Comparative Immunology*, 115, 103907. <https://doi.org/10.1016/j.dci.2020.103907>
- Han, X., Wang, L., Li, T., Zhang, J., Zhang, D., Li, J., Xia, Y., Liu, Y., & Tan, W. (2020). Beyond Blocking: Engineering RNAi-Mediated Targeted Immune Checkpoint Nanoblocker Enables T-Cell-Independent Cancer Treatment. *ACS Nano*, 14(12), 17524–17534. <https://doi.org/10.1021/acsnano.0c08022>
- He, C., Yue, H., Xu, L., Liu, Y., Song, Y., Tang, C., & Yin, C. (2020). siRNA release kinetics from polymeric nanoparticles correlate with RNAi efficiency and inflammation therapy via oral delivery. *Acta Biomaterialia*, 103, 213–222. <https://doi.org/10.1016/j.actbio.2019.12.005>
- Hung, Y.-H., & Slotkin, R. K. (2021). The initiation of RNA interference (RNAi) in plants. *Current Opinion in Plant Biology*, 61, 102014. <https://doi.org/10.1016/j.pbi.2021.102014>
- Jiang, T., Qiao, Y., Ruan, W., Zhang, D., Yang, Q., Wang, G., Chen, Q., Zhu, F., Yin, J., Zou, Y., Qian, R., Zheng, M., & Shi, B. (2021). Cation-Free siRNA Micelles as Effective Drug Delivery Platform and Potent RNAi Nanomedicines for Glioblastoma Therapy. *Advanced Materials*, 33(45), 2104779. <https://doi.org/10.1002/adma.202104779>
- Jin, Y., Zhao, J.-H., & Guo, H.-S. (2021). Recent advances in understanding plant antiviral RNAi and viral suppressors of RNAi. *Current Opinion in Virology*, 46, 65–72. <https://doi.org/10.1016/j.coviro.2020.12.001>
- Kara, G., Calin, G. A., & Ozpolat, B. (2022). RNAi-based therapeutics and tumor targeted delivery in cancer. *Advanced Drug Delivery Reviews*, 182, 114113. <https://doi.org/10.1016/j.addr.2022.114113>
- Kelleher, A. D., Cortez-Jugo, C., Cavalieri, F., Qu, Y., Glanville, A. R., Caruso, F., Symonds, G., & Ahlenstiel, C. L. (2020). RNAi therapeutics: An antiviral strategy for human infections. *Current Opinion in Pharmacology*, 54, 121–129. <https://doi.org/10.1016/j.coph.2020.09.011>
- Kim, D. S., & Zhang, J. (2023). Strategies to improve the efficiency of RNAi-mediated crop protection for pest control. *Entomologia Generalis*, 43(1), 5–19. <https://doi.org/10.1127/entomologia/2022/1638>
- Kimura, Y., Shu, Z., Ito, M., Abe, N., Nakamoto, K., Tomoike, F., Shuto, S., Ito, Y., & Abe, H. (2020). Intracellular build-up RNAi with single-strand circular RNAs as siRNA precursors. *Chemical Communications*, 56(3), 466–469. <https://doi.org/10.1039/C9CC04872C>
- Kolge, H., Kadam, K., Galande, S., Lanjekar, V., & Ghormade, V. (2021). New Frontiers in Pest Control: Chitosan Nanoparticles-Shielded dsRNA as an Effective Topical RNAi Spray for Gram Podborer Biocontrol. *ACS Applied Bio Materials*, 4(6), 5145–5157. <https://doi.org/10.1021/acsabm.1c00349>
- Kyre, B. R., Bentz, B. J., & Rieske, L. K. (2020). Susceptibility of mountain pine beetle (*Dendroctonus ponderosae* Hopkins) to gene silencing through RNAi provides potential as a novel management tool. *Forest Ecology and Management*, 473, 118322. <https://doi.org/10.1016/j.foreco.2020.118322>
- Laird, N. Z., Acri, T. M., Tingle, K., & Salem, A. K. (2021). Gene- and RNAi-activated scaffolds for bone tissue engineering: Current progress and future directions. *Advanced Drug Delivery Reviews*, 174, 613–627. <https://doi.org/10.1016/j.addr.2021.05.009>
- Laisney, J., Gurusamy, D., Baddar, Z. E., Palli, S. R., & Unrine, J. M. (2020). RNAi in *Spodoptera frugiperda* Sf9 Cells via Nanomaterial Mediated Delivery of dsRNA: A Comparison of Poly- L -arginine Polyplexes and Poly- L -arginine-Functionalized Au

- Nanoparticles. *ACS Applied Materials & Interfaces*, 12(23), 25645–25657. <https://doi.org/10.1021/acsami.0c06234>
- Lee, S., Kim, S., Koo, D.-J., Yu, J., Cho, H., Lee, H., Song, J. M., Kim, S.-Y., Min, D.-H., & Jeon, N. L. (2021). 3D Microfluidic Platform and Tumor Vascular Mapping for Evaluating Anti-Angiogenic RNAi-Based Nanomedicine. *ACS Nano*, 15(1), 338–350. <https://doi.org/10.1021/acsnano.0c05110>
- Li, K., Zhang, Y., Hussain, A., Weng, Y., & Huang, Y. (2021). Progress of Photodynamic and RNAi Combination Therapy in Cancer Treatment. *ACS Biomaterials Science & Engineering*, 7(9), 4420–4429. <https://doi.org/10.1021/acsbiomaterials.1c00765>
- Li, Q., Lv, X., Tang, C., & Yin, C. (2022). Co-delivery of doxorubicin and CRISPR/Cas9 or RNAi-expressing plasmid by chitosan-based nanoparticle for cancer therapy. *Carbohydrate Polymers*, 287, 119315. <https://doi.org/10.1016/j.carbpol.2022.119315>
- Liu, X., Xie, X., Zheng, C., Wei, L., Li, X., Jin, Y., Zhang, G., Jiang, C.-J., & Liang, Z. (2022). RNAi-mediated suppression of the abscisic acid catabolism gene OsABA8ox1 increases abscisic acid content and tolerance to saline–alkaline stress in rice (*Oryza sativa* L.). *The Crop Journal*, 10(2), 354–367. <https://doi.org/10.1016/j.cj.2021.06.011>
- Liu, Z., Zhao, L., Huang, L., Qin, Y., Zhang, J., Zhang, J., & Yan, Q. (2020). Integration of RNA-seq and RNAi provides a novel insight into the immune responses of *Epinephelus coioides* to the *impB* gene of *Pseudomonas plecoglossicida*. *Fish & Shellfish Immunology*, 105, 135–143. <https://doi.org/10.1016/j.fsi.2020.06.023>
- Mujtaba, M., Wang, D., Carvalho, L. B., Oliveira, J. L., Espirito Santo Pereira, A. D., Sharif, R., Jogaiah, S., Paidi, M. K., Wang, L., Ali, Q., & Fraceto, L. F. (2021). Nanocarrier-Mediated Delivery of miRNA, RNAi, and CRISPR-Cas for Plant Protection: Current Trends and Future Directions. *ACS Agricultural Science & Technology*, 1(5), 417–435. <https://doi.org/10.1021/acsaagscitech.1c00146>
- Obici, L., Berk, J. L., González-Duarte, A., Coelho, T., Gillmore, J., Schmidt, H. H.-J., Schilling, M., Yamashita, T., Labeyrie, C., Brannagan, T. H., Ajroud-Driss, S., Gorevic, P., Kristen, A. V., Franklin, J., Chen, J., Sweetser, M. T., Wang, J. J., & Adams, D. (2020). Quality of life outcomes in APOLLO, the phase 3 trial of the RNAi therapeutic patisiran in patients with hereditary transthyretin-mediated amyloidosis. *Amyloid*, 27(3), 153–162. <https://doi.org/10.1080/13506129.2020.1730790>
- Riga, M., Denecke, S., Livadaras, I., Geibel, S., Nauen, R., & Vontas, J. (2020). Development of efficient RNAi in *Nezara viridula* for use in insecticide target discovery. *Archives of Insect Biochemistry and Physiology*, 103(3), e21650. <https://doi.org/10.1002/arch.21650>
- Saify Nabiabad, H., Amini, M., & Demirdas, S. (2022). Specific delivering of RNAi using Spike's aptamer-functionalized lipid nanoparticles for targeting SARS-CoV-2: A strong anti-Covid drug in a clinical case study. *Chemical Biology & Drug Design*, 99(2), 233–246. <https://doi.org/10.1111/cbdd.13978>
- Schwartz-Orbach, L., Zhang, C., Sidoli, S., Amin, R., Kaur, D., Zhebrun, A., Ni, J., & Gu, S. G. (2020). *Caenorhabditis elegans* nuclear RNAi factor SET-32 deposits the transgenerational histone modification, H3K23me3. *eLife*, 9, e54309. <https://doi.org/10.7554/eLife.54309>
- Šečić, E., & Kogel, K.-H. (2021). Requirements for fungal uptake of dsRNA and gene silencing in RNAi-based crop protection strategies. *Current Opinion in Biotechnology*, 70, 136–142. <https://doi.org/10.1016/j.copbio.2021.04.001>
- Uddin, N., Binzel, D. W., Shu, D., Fu, T.-M., & Guo, P. (2023). Targeted delivery of RNAi to cancer cells using RNA-ligand displaying exosome. *Acta Pharmaceutica Sinica B*, 13(4), 1383–1399. <https://doi.org/10.1016/j.apsb.2022.11.019>
- Wang, J., Chen, G., Liu, N., Han, X., Zhao, F., Zhang, L., & Chen, P. (2022). Strategies for improving the safety and RNAi efficacy of noncovalent peptide/siRNA nanocomplexes.

- Advances in Colloid and Interface Science*, 302, 102638. <https://doi.org/10.1016/j.cis.2022.102638>
- Wiltshire, R. M., & Duman-Scheel, M. (2020). Advances in oral RNAi for disease vector mosquito research and control. *Current Opinion in Insect Science*, 40, 18–23. <https://doi.org/10.1016/j.cois.2020.05.002>
- Xin, G., Wang, F., Zhao, L., Qin, Y., Huang, L., & Yan, Q. (2020). Integration of RNA-seq and RNAi provides a novel insight into the effect of pvdE gene to the pathogenic of *Pseudomonas plecoglossicida* and on the immune responses of orange-spotted grouper (*Epinephelus coioides*). *Aquaculture*, 529, 735695. <https://doi.org/10.1016/j.aquaculture.2020.735695>
- Xu, L., Faruqu, F. N., Lim, Y. M., Lim, K. Y., Liam-Or, R., Walters, A. A., Lavender, P., Fear, D., Wells, C. M., Tzu-Wen Wang, J., & Al-Jamal, K. T. (2021). Exosome-mediated RNAi of PAK4 prolongs survival of pancreatic cancer mouse model after loco-regional treatment. *Biomaterials*, 264, 120369. <https://doi.org/10.1016/j.biomaterials.2020.120369>
- Yan, S., Ren, B., Zeng, B., & Shen, J. (2020). Improving RNAi Efficiency for Pest Control in Crop Species. *BioTechniques*, 68(5), 283–290. <https://doi.org/10.2144/btn-2019-0171>
- Yue, D., Cai, X., Fan, M., Zhu, J., Tian, J., Wu, L., Jiang, Q., & Gu, Z. (2021). An Alternating Irradiation Strategy-Driven Combination Therapy of PDT and RNAi for Highly Efficient Inhibition of Tumor Growth and Metastasis. *Advanced Healthcare Materials*, 10(8), 2001850. <https://doi.org/10.1002/adhm.202001850>
- Zhang, D., Sun, Y., Wang, S., Zou, Y., Zheng, M., & Shi, B. (2022). Brain-Targeting Metastatic Tumor Cell Membrane Cloaked Biomimetic Nanomedicines Mediate Potent Chemodynamic and RNAi Combinational Therapy of Glioblastoma. *Advanced Functional Materials*, 32(51), 2209239. <https://doi.org/10.1002/adfm.202209239>
- Zhang, K., Wei, J., Huff Hartz, K. E., Lydy, M. J., Moon, T. S., Sander, M., & Parker, K. M. (2020). Analysis of RNA Interference (RNAi) Biopesticides: Double-Stranded RNA (dsRNA) Extraction from Agricultural Soils and Quantification by RT-qPCR. *Environmental Science & Technology*, 54(8), 4893–4902. <https://doi.org/10.1021/acs.est.9b07781>
- Zhang, W., Han, B., Lai, X., Xiao, C., Xu, S., Meng, X., Li, Z., Meng, J., Wen, T., Yang, X., Liu, J., & Xu, H. (2020). Stiffness of cationized gelatin nanoparticles is a key factor determining RNAi efficiency in myeloid leukemia cells. *Chemical Communications*, 56(8), 1255–1258. <https://doi.org/10.1039/C9CC09068A>

---

**Copyright Holder :**

© Dito Anurogo et al. (2025).

**First Publication Right :**

© Journal of Biomedical and Techno Nanomaterials

**This article is under:**

