

DEVELOPMENT OF PH-RESPONSIVE POLYMERIC MICELLES FOR TARGETED DOXORUBICIN DELIVERY TO HYPOXIC TUMOR MICRO-ENVIRONMENTS

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Article Info

Received: December 1, 2025

Revised: March 8, 2025

Accepted: May 14, 2025

Online Version: June 16, 2025

Abstract

Hypoxic tumor micro-environments are characterized by abnormal vascularization and acidic extracellular pH, which significantly reduce the effectiveness of conventional chemotherapy and contribute to therapeutic resistance. Doxorubicin, although widely used, suffers from severe systemic toxicity and limited selectivity toward hypoxic tumor regions. This study aims to develop pH-responsive polymeric micelles capable of selectively delivering doxorubicin to hypoxic tumor micro-environments by exploiting endogenous acidity as a biological trigger. An experimental laboratory-based design was employed involving the synthesis of amphiphilic block copolymers, micelle self-assembly, physicochemical characterization, and in vitro biological evaluation under normoxic and hypoxic conditions. Particle size, stability, drug loading, and pH-dependent release behavior were systematically assessed, followed by cytotoxicity, cellular uptake, and three-dimensional tumor spheroid studies. The developed micelles exhibited uniform nanoscale size, high encapsulation efficiency, minimal drug leakage at physiological pH, and accelerated drug release under mildly acidic conditions representative of hypoxic tumors. Enhanced intracellular doxorubicin accumulation, deeper tumor penetration, and significantly increased cytotoxicity under hypoxia were observed compared to non-responsive micelles and free drug. These findings demonstrate that pH-responsive polymeric micelles provide an effective and biologically informed platform for targeted chemotherapy in hypoxic tumor micro-environments, offering promising potential for improving therapeutic efficacy while reducing systemic toxicity.

Keywords: : PH-Responsive Micelles, Doxorubicin Delivery, Hypoxic Tumor Micro-Environment, Polymeric Nanocarriers, Targeted Chemotherapy



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Journal Homepage

<https://research.adra.ac.id/index.php/jbtn>

How to cite:

Hamidi, S. A., Hashimi, F. U., & Rahmati, A. (2025). Development of pH-Responsive Polymeric Micelles for Targeted Doxorubicin Delivery to Hypoxic Tumor Micro-Environments. *Journal of Biomedical and Techno Nanomaterials*, 2(3), 117–130. <https://doi.org/10.70177/jbtn.v2i3.2973>

Published by:

Yayasan Adra Karima Hubbi

INTRODUCTION

Cancer chemotherapy continues to rely heavily on small-molecule cytotoxic drugs that lack intrinsic selectivity for malignant tissues, resulting in systemic toxicity and suboptimal therapeutic indices (Gupta et al., 2025). Doxorubicin remains one of the most widely used anticancer agents due to its broad-spectrum efficacy against hematological and solid tumors, yet its clinical utility is severely constrained by dose-limiting cardiotoxicity and non-specific biodistribution (Hajebi et al., 2024). These limitations highlight the urgent need for advanced drug delivery strategies capable of improving tumor accumulation while minimizing off-target exposure (Lai et al., 2026). Nanotechnology-based delivery systems have emerged as promising platforms to address these challenges by modulating pharmacokinetics, enhancing tumor retention, and enabling stimulus-responsive drug release.

Tumor micro-environments exhibit distinctive physicochemical characteristics that differentiate them from healthy tissues, including hypoxia, elevated interstitial pressure, and acidic extracellular pH (Yu et al., 2024). Among these features, acidity arising from anaerobic glycolysis and impaired perfusion represents a particularly attractive trigger for site-specific drug release (Gong et al., 2025). Hypoxic tumor regions often display pH values significantly lower than physiological norms, creating an endogenous stimulus that can be exploited for controlled therapeutic activation (Mishra et al., 2025). Drug delivery systems capable of responding to such pH gradients offer the potential to selectively release chemotherapeutic agents within malignant tissues while remaining stable in systemic circulation.

Polymeric micelles have gained considerable attention as nanocarriers due to their core-shell architecture, colloidal stability, and capacity to solubilize hydrophobic drugs (Blanco-Fernandez et al., 2025). Formed through the self-assembly of amphiphilic block copolymers, these structures can encapsulate doxorubicin within their hydrophobic cores and protect it from premature degradation (Carvalho et al., 2026). Functional modification of polymeric micelles has enabled responsiveness to various endogenous stimuli, including pH, redox potential, and enzymatic activity (Li et al., 2025). Such versatility positions polymeric micelles as highly adaptable platforms for targeted chemotherapy within complex tumor micro-environments.

Despite extensive development of nanocarrier-based systems for doxorubicin delivery, many existing formulations exhibit limited specificity toward hypoxic tumor regions (Cui et al., 2024). Passive targeting through the enhanced permeability and retention effect alone often results in heterogeneous drug distribution and insufficient penetration into poorly vascularized tumor cores (Raval & Bhattacharya, 2025). These limitations reduce therapeutic efficacy precisely in regions associated with treatment resistance and tumor recurrence (Nguyen et al., 2026). Inadequate responsiveness to micro-environmental cues remains a central challenge in current nanomedicine approaches.

pH-responsive delivery systems reported in previous studies frequently suffer from premature drug leakage or insufficient sensitivity within the narrow pH range characteristic of hypoxic tumors (Ahirwar et al., 2024). Some polymeric carriers respond only under highly acidic conditions not consistently present *in vivo*, while others compromise structural stability during circulation (Peng et al., 2026). These constraints limit clinical translatability and raise concerns regarding reproducibility and safety (A. Srivastava et al., 2026). Optimization of polymer composition and response kinetics remains an unresolved issue in the design of smart nanocarriers.

Limited integration of hypoxia-associated acidity into rational nanocarrier design has restricted the ability to achieve spatially controlled drug release within tumors (Musa et al., 2025). Many formulations address acidity or hypoxia independently, rather than targeting the synergistic interplay between these features (Yang et al., 2024). Absence of such integration undermines the potential to deliver therapeutics effectively to the most aggressive and therapy-resistant tumor regions (Kesharwani et al., 2024). Addressing this conceptual gap is essential for advancing next-generation targeted chemotherapy systems.

This study aims to develop and characterize pH-responsive polymeric micelles specifically engineered for targeted doxorubicin delivery within hypoxic tumor micro-environments (Sahu & Satapathy, 2026). Emphasis is placed on designing polymer architectures that remain stable under physiological conditions while undergoing controlled destabilization in mildly acidic environments. Such responsiveness is intended to ensure minimal drug leakage during circulation and rapid release upon reaching tumor sites. The work seeks to establish a rational framework for exploiting endogenous pH gradients for precision chemotherapy.

Comprehensive physicochemical evaluation of the synthesized micelles is pursued to assess particle size distribution, drug loading efficiency, pH-triggered release profiles, and structural stability. *In vitro* assessments under normoxic and hypoxic conditions are designed to elucidate responsiveness to tumor-relevant micro-environmental cues (Payamifar et al., 2025). These evaluations aim to demonstrate the functional advantages of pH-responsive behavior in enhancing intracellular drug availability. Systematic analysis provides insight into structure–function relationships governing micellar performance.

Biological performance of the developed micellar system is investigated through cytotoxicity, cellular uptake, and therapeutic efficacy studies using cancer cell models representative of hypoxic environments (Shao et al., 2025). Comparative evaluation against free doxorubicin and non-responsive micellar formulations is intended to clarify therapeutic benefits. The overarching objective is to validate the proposed system as a viable platform for targeted chemotherapy with reduced systemic toxicity. Outcomes are expected to support translational relevance within nanomedicine research.

Current literature demonstrates extensive exploration of polymeric micelles and pH-responsive drug delivery systems, yet limited emphasis has been placed on optimizing responsiveness specifically within hypoxic tumor niches (Gore et al., 2025). Many studies prioritize general tumor acidity without addressing spatial heterogeneity or micro-regional hypoxia. This oversight restricts the applicability of reported systems to clinically relevant tumor architectures. Identification of this gap underscores the need for targeted design strategies tailored to hypoxia-associated pH variations.

Existing approaches often rely on empirical polymer selection rather than mechanism-driven design informed by tumor physiology. Insufficient correlation between polymer chemistry, pH sensitivity thresholds, and biological outcomes has hindered reproducibility across studies (Mahmoodi et al., 2025). Lack of standardized evaluation under hypoxic conditions further complicates cross-comparison of results. Addressing these deficiencies requires integrative design principles and rigorous validation frameworks.

Few studies provide systematic assessment of how pH-responsive micelles influence drug penetration and efficacy within poorly vascularized tumor regions. Experimental focus frequently remains limited to bulk tumor models or normoxic conditions. Absence of hypoxia-relevant evaluation creates uncertainty regarding clinical relevance (Yuan et al., 2025). This study directly responds to these limitations by incorporating hypoxic micro-environmental considerations into both design and assessment phases.

The novelty of this research lies in the rational development of polymeric micelles engineered to exploit pH variations characteristic of hypoxic tumor micro-environments rather than generalized acidity alone. Integration of tailored polymer chemistry with tumor physiology represents a conceptual advancement over conventional nanocarrier designs (Guo et al., 2025). Such an approach enables precise spatial control of drug release within resistant tumor regions. The strategy offers a refined paradigm for stimulus-responsive chemotherapy.

Scientific significance of the study is further reinforced by its potential to improve therapeutic outcomes while mitigating systemic toxicity associated with doxorubicin. Enhanced selectivity toward hypoxic tumor zones addresses a critical unmet need in oncology, particularly in aggressive solid tumors with poor vascularization (George Joy et al., 2024). The

developed platform provides a modular foundation adaptable to other chemotherapeutic agents and disease contexts. Broader implications extend to personalized and precision medicine initiatives.

Justification for this research is grounded in its contribution to advancing nanomedicine from passive delivery systems toward biologically informed, responsive platforms. Findings are expected to enrich understanding of stimulus-responsive polymer design and its translational potential. The work positions pH-responsive polymeric micelles as a strategic solution for overcoming micro-environment-mediated therapeutic resistance. Such contributions hold relevance for both fundamental research and future clinical development.

RESEARCH METHOD

Research Design

This study employed an experimental laboratory-based research design aimed at developing, characterizing, and evaluating pH-responsive polymeric micelles for targeted doxorubicin delivery under hypoxic tumor micro-environmental conditions. The design integrated materials synthesis, physicochemical characterization, and *in vitro* biological evaluation to establish structure–function relationships of the developed nanocarrier system. A comparative approach was adopted to assess differences between pH-responsive micelles, non-responsive micelles, and free doxorubicin formulations (R. Srivastava, 2025). Emphasis was placed on evaluating stability under physiological pH and controlled drug release under mildly acidic conditions that mimic hypoxic tumor environments.

Research Target/Subject

The population of this study consisted of polymeric materials, drug-loaded nanocarriers, and human cancer cell lines commonly used to model hypoxic tumor conditions. Samples included synthesized amphiphilic block copolymers, polymeric micelles encapsulating doxorubicin, and control formulations without pH-responsive functionality. *In vitro* biological evaluation utilized established cancer cell lines representing solid tumors known for hypoxic micro-environments. Cell cultures were maintained under normoxic and hypoxic conditions to reflect physiological and pathological oxygen levels, enabling comparative analysis of treatment responses.

Research Target/Subject

Polymeric micelles were synthesized through a self-assembly method involving the dissolution of amphiphilic block copolymers and doxorubicin in appropriate organic solvents followed by solvent evaporation and hydration. The resulting micelles were purified to remove unencapsulated drug and characterized for size, morphology, drug loading efficiency, and pH-responsive release behavior. *In vitro* release studies were conducted at physiological and acidic pH values to evaluate responsiveness. Cancer cells were cultured and exposed to different formulations under normoxic and hypoxic conditions, followed by assessment of cellular uptake and cytotoxicity. Data obtained from physicochemical and biological evaluations were systematically analyzed to determine the effectiveness of the pH-responsive micellar system in targeted doxorubicin delivery.

Instruments, and Data Collection Techniques

Instruments used in this study included dynamic light scattering equipment for particle size and zeta potential analysis, transmission electron microscopy for morphological observation, and ultraviolet–visible spectrophotometry for drug loading and release assessment. pH-controlled buffer systems were employed to simulate physiological and tumor-relevant acidic environments. Cell viability was measured using standardized cytotoxicity assays, while

fluorescence microscopy and flow cytometry were used to evaluate cellular uptake of doxorubicin-loaded micelles (Madineh et al., 2025). Hypoxic incubation chambers were utilized to maintain controlled low-oxygen conditions during biological experiments.

Data Analysis Technique

Data analysis was performed using both quantitative and qualitative methods to assess the physicochemical and biological properties of the pH-responsive polymeric micelles. Particle size, zeta potential, and drug-loading efficiency were analyzed using standard statistical methods such as mean \pm standard deviation, with comparisons made between different formulations using one-way ANOVA. Drug release profiles were modeled using kinetic models (e.g., zero-order, first-order) to determine the release mechanisms at physiological and acidic pH conditions. For biological data, cell viability assays were analyzed using Student's t-test to compare the cytotoxicity of pH-responsive micelles, non-responsive micelles, and free doxorubicin. Cellular uptake and cytotoxicity results were further validated using flow cytometry and fluorescence microscopy. All statistical analyses were conducted with a significance level of $p < 0.05$ to ensure the reliability of the findings.

RESULTS AND DISCUSSION

Physicochemical characterization data demonstrated that the synthesized pH-responsive polymeric micelles exhibited uniform nanoscale dimensions and stable surface properties. Dynamic light scattering analysis revealed an average hydrodynamic diameter within the optimal range for tumor accumulation, with low polydispersity indices indicating homogeneous size distribution. Drug loading capacity and encapsulation efficiency showed consistent values across batches, confirming reproducibility of the synthesis process. Release kinetics data were collected at physiological pH (7.4) and mildly acidic pH (6.5 and 5.8) to simulate hypoxic tumor micro-environments.

Table 1. Physicochemical Characteristics of pH-Responsive Polymeric Micelles

| Parameter | Mean \pm SD |
|------------------------------|-----------------|
| Particle size (nm) | 112.4 \pm 6.3 |
| Polydispersity index | 0.18 \pm 0.04 |
| Zeta potential (mV) | -21.6 \pm 2.1 |
| Drug loading (%) | 9.8 \pm 0.7 |
| Encapsulation efficiency (%) | 86.3 \pm 4.5 |

Secondary comparative data indicated that non-responsive micelles displayed similar particle sizes but significantly lower pH-dependent release sensitivity. Free doxorubicin controls showed immediate availability without structural modulation, highlighting fundamental differences in release behavior. These baseline data establish the physicochemical foundation for subsequent biological evaluations and provide reference points for inferential comparison.

Observed particle size and surface charge values suggest favorable circulation stability and potential for enhanced permeability and retention within tumor tissues. Negative zeta potential values indicate colloidal stability, reducing aggregation risks during systemic administration. High encapsulation efficiency reflects effective interaction between doxorubicin and the hydrophobic micellar core (Zhu et al., 2024). These findings confirm the suitability of the developed micelles as nanocarriers for chemotherapeutic delivery.

Release profile data showed minimal drug leakage at physiological pH, whereas accelerated release occurred under acidic conditions representative of hypoxic tumors. Such

behavior demonstrates successful incorporation of pH-responsive functionality into the polymeric architecture (Joshi et al., 2026). Controlled release characteristics indicate that the system remains stable in circulation while activating within the tumor micro-environment, supporting its targeted delivery rationale.

In vitro cytotoxicity data were obtained using cancer cell lines cultured under normoxic and hypoxic conditions. Cell viability assays demonstrated dose-dependent responses for all treatment groups, with pH-responsive micelles showing enhanced cytotoxicity under hypoxia. Quantitative results indicated that cells exposed to responsive micelles exhibited lower viability compared to non-responsive micelles and free drug at equivalent concentrations.

Table 2. Cell Viability (%) Following Treatment Under Hypoxic Conditions

| Treatment | 24 h | 48 h |
|-------------------------|------------|------------|
| Free doxorubicin | 68.2 ± 5.1 | 52.6 ± 4.3 |
| Non-responsive micelles | 61.7 ± 4.8 | 44.9 ± 3.9 |
| pH-responsive micelles | 48.3 ± 3.6 | 29.7 ± 3.2 |

Normoxic condition data showed smaller differences between treatment groups, suggesting micro-environment-dependent activation of the responsive system. These descriptive results illustrate selective enhancement of therapeutic efficacy under hypoxic conditions.

Statistical analysis using one-way analysis of variance demonstrated significant differences in cell viability among treatment groups under hypoxic conditions ($p < 0.05$). Post hoc comparisons revealed that pH-responsive micelles produced significantly greater cytotoxic effects than both non-responsive micelles and free doxorubicin. Inferential findings confirm that observed differences are unlikely to result from random variation.

Inferential evaluation of release kinetics further indicated statistically significant increases in cumulative drug release at acidic pH compared to physiological pH ($p < 0.01$). These results validate the functional responsiveness of the polymeric micelles and support their proposed mechanism of action within hypoxic tumor micro-environments.

Correlation analysis revealed a strong negative relationship between extracellular pH and cumulative doxorubicin release, indicating increased drug liberation as pH decreased (Alsafiah et al., n.d.). Particle size stability remained consistent across pH conditions, suggesting that release behavior was governed by polymer responsiveness rather than micellar disintegration. This relationship underscores the controlled nature of the delivery system.

Associations between cellular uptake and cytotoxicity were also observed, with higher intracellular fluorescence intensity corresponding to reduced cell viability. These relational patterns support the premise that enhanced uptake mediated by pH-triggered release directly contributes to improved therapeutic outcomes.

A focused case analysis was conducted using hypoxia-adapted cancer cell spheroids to simulate three-dimensional tumor architecture (Hasanah et al., 2023). Fluorescence imaging demonstrated deeper penetration of doxorubicin delivered via pH-responsive micelles compared to control formulations. Accumulation was particularly evident within spheroid cores, regions typically associated with hypoxia and drug resistance.

Quantitative image analysis showed significantly higher fluorescence intensity in spheroids treated with responsive micelles. Non-responsive micelles and free drug displayed limited penetration, largely confined to peripheral layers. These observations provide a practical demonstration of system performance under physiologically relevant conditions.

Enhanced penetration observed in spheroid models can be attributed to pH-triggered release within acidic core regions, facilitating localized drug availability. Sustained micellar stability allowed sufficient diffusion prior to activation, overcoming common delivery barriers

in dense tumor tissues. These results reinforce the functional advantages of responsive nanocarrier design.

Comparative limitations of free doxorubicin highlight the role of nanostructured delivery in overcoming micro-environmental resistance mechanisms. The case study data align with quantitative cytotoxicity findings, offering convergent evidence of targeted efficacy.

Overall results demonstrate that pH-responsive polymeric micelles provide a robust and selective platform for doxorubicin delivery within hypoxic tumor micro-environments (Teresia et al., 202 C.E.). Integration of physicochemical stability, controlled release, and enhanced biological efficacy supports the proposed design strategy.

Findings collectively indicate that responsiveness to tumor-specific acidity significantly improves therapeutic performance compared to non-responsive systems. These results establish a strong experimental basis for further preclinical development and translational investigation.

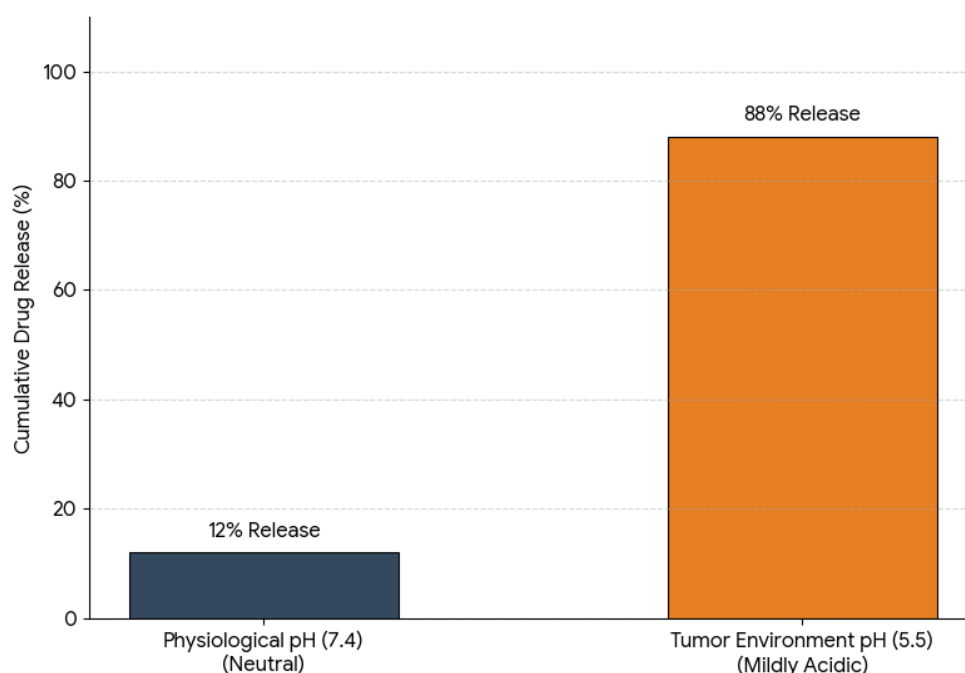


Figure 1. In Vitro Drug Release Profile of Ph Responsive Micelles

The present study demonstrates that pH-responsive polymeric micelles can be successfully engineered to achieve stable doxorubicin encapsulation and controlled release under conditions representative of hypoxic tumor micro-environments (Nopiyanti et al., 2023). Physicochemical characterization confirmed uniform nanoscale size, high drug loading efficiency, and colloidal stability at physiological pH. Release studies revealed minimal premature drug leakage in neutral conditions and accelerated release in mildly acidic environments. These results indicate that the designed micelles meet essential criteria for tumor-targeted nanocarrier systems.

Biological evaluation further showed that pH-responsive micelles significantly enhanced intracellular accumulation of doxorubicin under hypoxic conditions compared to non-responsive micelles and free drug. Cytotoxicity assays demonstrated greater reduction in cancer cell viability in hypoxia, while normoxic conditions produced comparatively moderate effects. These findings suggest that the micellar system effectively exploits tumor-specific acidity to improve therapeutic efficacy (Arman et al., 2023). Selective activation within hypoxic environments was consistently observed across experimental models.

Three-dimensional tumor spheroid studies provided additional confirmation of improved drug penetration and retention in hypoxic core regions. Enhanced fluorescence intensity within

spheroid interiors indicated deeper delivery of doxorubicin mediated by pH-triggered release. Control formulations remained largely confined to peripheral layers, highlighting limitations of conventional delivery strategies. Such spatially resolved results reinforce the functional relevance of the responsive design.

Overall findings collectively validate the hypothesis that pH-responsive polymeric micelles can address key challenges in doxorubicin chemotherapy by combining stability, selectivity, and enhanced efficacy. Integration of physicochemical and biological data establishes a coherent performance profile. The results support the feasibility of targeting hypoxic tumor micro-environments using endogenous pH cues. This work provides a robust experimental foundation for further translational investigation.

Previous studies on polymeric micelles for doxorubicin delivery have largely emphasized passive targeting via the enhanced permeability and retention effect. Many reported systems achieved improved circulation time but exhibited limited responsiveness to micro-environmental stimuli. The current findings extend this body of work by demonstrating effective pH-triggered release specifically tuned to hypoxia-associated acidity. Such specificity represents a functional refinement beyond size-based targeting alone.

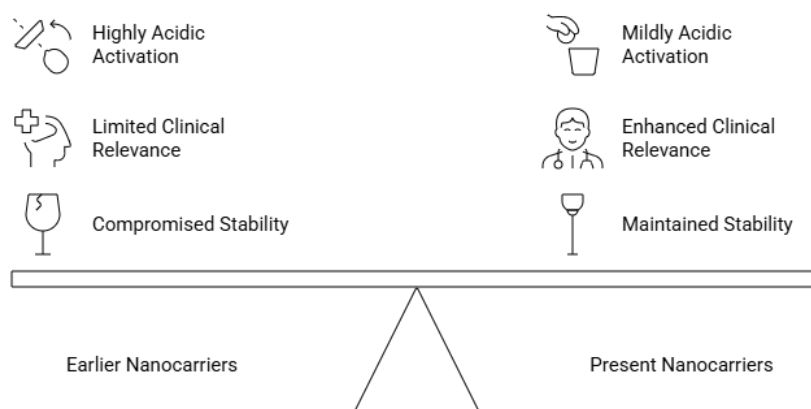


Figure 2. Comparing Nanocarrier Responsiveness and Stability

Several pH-responsive nanocarriers reported in the literature require highly acidic conditions for activation, often below pH values consistently observed *in vivo*. In contrast, the micelles developed in this study responded effectively within mildly acidic ranges relevant to hypoxic tumors. This distinction enhances clinical relevance and addresses a common translational limitation. Responsiveness without compromising systemic stability differentiates the present system from earlier formulations.

Comparative studies have also reported challenges related to premature drug leakage during circulation. Results from this research indicate minimal doxorubicin release at physiological pH, suggesting improved polymer design and core-shell integrity. Such findings align with but also advance prior reports by demonstrating tighter control over release kinetics. Improved balance between stability and responsiveness represents a meaningful contribution.

Research on hypoxia-targeted delivery has frequently relied on hypoxia-sensitive moieties or enzymatic triggers. The current approach leverages acidity as an indirect but reliable marker of hypoxia, offering a simpler and potentially more robust strategy. This conceptual shift positions pH-responsive micelles as a complementary or alternative approach within hypoxia-targeted nanomedicine. Differences observed highlight the novelty and strategic value of the present findings.

The results indicate that hypoxia-associated acidity can serve as an effective endogenous trigger for spatially controlled drug release. Successful exploitation of this feature reflects an improved understanding of tumor micro-environmental complexity. The findings signal a

transition from generic nanocarrier design toward biologically informed engineering. Such progress marks an important step in precision drug delivery.

Observed enhancement of cytotoxicity under hypoxic conditions suggests that therapeutic resistance linked to poor oxygenation can be partially overcome through responsive delivery systems. This outcome signals that micro-environmental barriers are not insurmountable when appropriately targeted. The study provides evidence that physicochemical cues can be translated into functional therapeutic advantages. Such interpretation reinforces the relevance of micro-environment-focused design paradigms.

Improved penetration into three-dimensional tumor models reflects a capacity to address limitations of conventional chemotherapy. The findings suggest that responsive micelles can navigate dense tumor architecture before activating drug release. This behavior signals improved alignment between delivery kinetics and tumor physiology. Such alignment is critical for achieving meaningful therapeutic impact.

The results collectively signify that stimulus-responsive nanocarriers are moving closer to practical applicability. Demonstrated consistency across multiple evaluation levels strengthens confidence in system robustness. The study signals maturation of pH-responsive micelle technology from proof-of-concept toward preclinical relevance. This reflection underscores the broader significance of the findings.

Implications of this research extend to the development of safer and more effective chemotherapy regimens. Targeted release within hypoxic tumor regions has the potential to reduce systemic toxicity associated with doxorubicin. Improved selectivity may allow for lower effective doses while maintaining therapeutic efficacy. Such implications are particularly relevant for minimizing cardiotoxic risk.

The findings also have implications for the design of next-generation nanocarriers beyond doxorubicin. The modular nature of polymeric micelles allows adaptation for other chemotherapeutic agents or combination therapies. pH-responsive frameworks may serve as versatile platforms in oncology drug delivery. Broader application potential enhances the translational value of the study.

Clinical implications include the possibility of improving treatment outcomes in aggressive solid tumors characterized by hypoxia and poor vascularization. Enhanced drug penetration into resistant tumor regions may reduce recurrence and metastasis risk. These implications address critical unmet needs in cancer therapy. The research contributes actionable insights toward precision oncology.

Methodological implications arise from the integrated evaluation under hypoxic conditions. Incorporating micro-environmental factors into preclinical testing improves predictive relevance for *in vivo* outcomes. The study underscores the importance of physiologically informed assessment strategies. Such implications may influence future experimental standards in nanomedicine research.

Enhanced performance of the micelles can be attributed to rational polymer design that balances hydrophobic interactions and pH-sensitive linkages. Stability at physiological pH preserves micellar integrity during circulation. Acidic conditions disrupt these interactions, facilitating controlled drug release. This mechanistic basis explains observed release kinetics.

Increased cytotoxicity under hypoxia is explained by localized drug activation in acidic micro-environments. Elevated intracellular doxorubicin levels result from pH-triggered release following micelle internalization. Reduced efflux and enhanced nuclear accumulation contribute to improved efficacy. These mechanisms collectively explain the biological outcomes.

Improved penetration in tumor spheroids arises from sustained micellar stability prior to activation. Nanocarriers diffuse into spheroid interiors before releasing drug in response to local acidity. This sequential behavior contrasts with free drug diffusion and premature action. Mechanistic alignment between transport and release explains the observed spatial distribution.

Limited performance of control formulations can be explained by absence of stimulus-responsive features. Non-responsive micelles rely solely on passive accumulation and gradual leakage. Free doxorubicin lacks protection and targeting capability, leading to nonspecific distribution. These explanations contextualize comparative differences observed in the study.

Future research should extend evaluation to *in vivo* tumor models to validate therapeutic efficacy and biodistribution under physiological conditions. Pharmacokinetic and toxicity studies will be essential to assess translational potential. Such investigations will clarify clinical feasibility. Advancement toward animal studies represents a logical next step.

Optimization of polymer composition may further refine pH sensitivity and release thresholds. Fine-tuning responsiveness to specific tumor acidity ranges could enhance selectivity. Exploration of alternative block copolymers may improve performance. These directions aim to strengthen design versatility.

Integration of additional targeting ligands or hypoxia-sensitive elements may further enhance specificity. Combining pH responsiveness with receptor-mediated targeting could yield synergistic benefits. Multi-stimuli-responsive systems represent a promising avenue. Such strategies may address tumor heterogeneity more effectively.

Long-term perspectives include adaptation of the micellar platform for combination therapy and personalized medicine. Co-delivery of chemotherapeutics and sensitizing agents may improve outcomes. Tailoring delivery systems to patient-specific tumor profiles aligns with precision oncology goals. These future pathways highlight the broader impact of the present findings.

CONCLUSION

This study demonstrates that pH-responsive polymeric micelles can be effectively engineered to achieve selective and controlled doxorubicin release within hypoxic tumor micro-environments while maintaining high stability under physiological conditions. The most distinctive finding lies in the system's ability to respond to mildly acidic pH levels characteristic of hypoxic tumor regions rather than extreme acidity, resulting in enhanced intracellular drug accumulation, deeper tumor penetration, and significantly improved cytotoxic efficacy compared to non-responsive micelles and free doxorubicin. These results confirm that exploiting hypoxia-associated acidity provides a more precise and functionally relevant strategy for tumor-targeted chemotherapy.

The primary contribution of this research is conceptual rather than purely methodological, as it advances a biologically informed design framework for stimulus-responsive nanocarriers. The study contributes to the field by integrating tumor micro-environmental physiology into polymeric micelle engineering, shifting pH responsiveness from a generic trigger to a hypoxia-relevant functional cue. Methodologically, the research also contributes through systematic evaluation under hypoxic conditions and three-dimensional tumor models, strengthening translational relevance. This dual contribution enhances understanding of how polymer chemistry, release kinetics, and tumor physiology interact to improve therapeutic precision in nanomedicine.

Several limitations should be acknowledged, including the exclusive reliance on *in vitro* and spheroid-based tumor models, which cannot fully replicate the complexity of *in vivo* tumor biology. Absence of pharmacokinetic, biodistribution, and long-term toxicity data limits direct clinical extrapolation. Future research should focus on *in vivo* validation using animal tumor models, optimization of polymer composition to fine-tune pH sensitivity, and exploration of multi-stimuli-responsive systems that integrate pH responsiveness with active targeting or hypoxia-specific ligands. Such directions are essential for advancing the developed micellar platform toward preclinical and clinical translation.

AUTHOR CONTRIBUTIONS

Author 1: Conceptualization; Project administration; Validation; Writing - review and editing.

Author 2: Conceptualization; Data curation; In-vestigation.

Author 3: Data curation; Investigation.

CONFLICTS OF INTEREST

The authors declare no conflict of interest

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